



NBSIR 81-2314

Migration of Low Molecular Weight Additives in Polymers

U.S. DEPARTMENT OF COMMERCE National Bureau of Standards Center for Materials Science Polymer Science and Standards Division Washington, DC 20234

Semi-Annual Report for the Period October 1, 1980 - March 31, 1981 Issued September 1981

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Prepared for

Bureau of Foods Food and Drug Administration Washington, DC 20201



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L. E. Smith, S. S. Chang, and G. A. Senich

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NATIONAL BUREAU OF STANDARDS, Ernest Ambler, Director



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Polymer Science and Standards Division

April 29, 1981

This is a progress report of continuing work for the internal uses of a sponsoring agency. The results are incomplete and the conclusions are necessarily tentative. Any or all of the material in this report may be modified before the final report is issued.



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Summary

In the first half of the FY81, several new areas were investigated. In testing and searching for oil-simulating solvents, we have studied the extraction behavior of (a) n-octanol, an extracting solvent used widely in biological and environmental studies, (b) HB307, a synthetic triglyceride mixture used widely in Europe as food-oil simulant and (c) 95% ethanol, to assess the effect of 5% water on the extractability of anhydrous ethanol. Antioxidant migration in two (ethylene-vinyl acetate) copolymers having vinyl acetate contents of 5% and 13% is being investigated for comparison with the polyolefin data. We are also beginning investigations of vinyl chloride migration from poly(vinylchloride). The following summarizes some preliminary conclusions from these studies.

n-Octanol vs. Ethanol

Diffusion coefficients of the three migrants from branched polyethylene into n-octanol are about the same or slightly greater than those into ethanol at both 30 and 60°C. However, for migration of BHT from linear polyethylene, the diffusion coefficients may be 2 to 4 times higher for extractions with n-octanol compared to ethanol.

HB307 vs. Ethanol or other Triglycerides

Diffusion coefficients of the three migrants mentioned above were measured from branched polyethylene into HB307, a synthetic triglyceride mixture, at 40 and 60°C. The diffusion coefficients are found to be about equal to those for extractions into ethanol or other triglycerides, as expected.

95% Ethanol vs. Anhydrous Ethanol

The diffusion coefficients into 95% ethanol are generally reduced by a factor of 2/3 to 1/2 of those into anhydrous ethanol. However, the amount extractable by 95% ethanol is more strongly governed by a change in the partition coefficient, as a result of reduced solubility of some of the migrants such as $n-C_{32}H_{66}$ in 95% ethanol.

BHT from Ethylene-Vinyl Acetate Copolymers

Diffusion coefficient measurements of BHT from (ethylene-5% vinyl acetate) copolymer are currently in progress. In n-heptane, the diffusivity of BHT in the copolymer is only slightly higher than in branched polyethylene. However, at 60°C the diffusion coefficients into ethanol and corn oil are about three times greater than those for branched polyethylene extractions.

I. Extraction of Polyethylene with Oil Simulating Solvents
A. Comparison of n-Octanol and Ethanol Extractions

n-Octanol has been used widely in biological and environmental studies as an extracting solvent. We have studied the migration behavior of n- $C_{18}H_{38}$, $n-C_{32}H_{66}$, and BHT from both linear and branched polyethylenes into n-octanol at 30 and 60°C. The results are compared with those from ethanol extractions in Table I and summarized in Figure 1.

n-Octanol generally behaves as a slightly accelerating solvent for all three migrants, differing from results for anhydrous ethanol by a factor of less than 1.3. Only in the case of BHT migration from LPE is a greater accelerating factor of 3 to 4 evident. The large increase in the diffusion coefficient in this case may be due to the increased mobility of degraded fragments from BHT in this particular sample. This speculation is impossible to confirm without performing a full analysis.

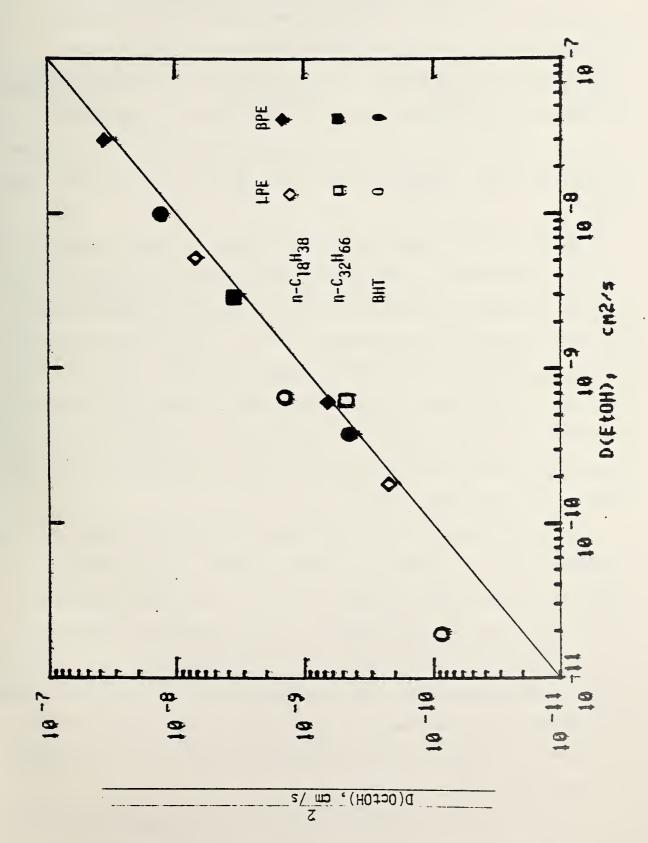
In the case of $n-C_{32}H_{66}$ migration, where partition coefficients are measureable in both ethanol and n-octanol extractions, the partition coefficients for n-octanol are nearly twice those of ethanol.

Thus, n-octanol is a slightly accelerating solvent when compared to ethanol and also allows a larger amount of oligomer to be extracted at equilibrium.

Table 1 Additive Migration into Ethanol and n-Octanol D, $\mu\text{m}^2\text{s}^{-1}$

			<u>Etha:</u>		<u>n-Octa</u>	
Sample Code	Polymer	Migrant	30°C	60°C	30°C	60°C
1,2	LPE	C18	0.018		0.022	
				0.52		0.70
7,9	BPE	C18	0.061		0.065	
				3.0		3.6
16	BPE	C32		0.29		0.35
19	LPE	C32		0.063		0.047
22	LPE	внт	0.0019		0.0087	
				0.047		0.14
25	BPE	BHT	0.038		0.045	
				1.0		1.3

Figure 1. Comparison of Diffusion Coefficients in n-Octanol and Ethanol



B. Comparison of Synthetic Triglyceride Mixture HB307 to Corn Oil

The synthetic triglyceride mixture HB307 has been used quite widely in some European laboratories as a food-oil simulant for extraction purposes. The particular sample of HB307 received by us differs slightly from the "typical" melting point of 28°C. It melts completely at 31-34°C as determined by differential scanning calorimetry. Because of the high melting point, extractions of the three migrants $(n-C_{18}H_{38}, n-C_{32}H_{66},$ and BHT) from branched polyethylene (sample codes 9, 16 and 25) with HB307 were performed at 40 and 60°C.

The extraction data for HB307 at 60°C are shown in Figure 2 together with the corresponding corn oil data obtained previously and normalized to the same thickness. The results for branched polyethylene extractions by both HB307 and corn oil are summarized in Table 2. It can be seen that for $n-C_{18}H_{38}$ the diffusion coefficients into HB307 are slightly lower than those into corn oil. For both $n-C_{32}H_{66}$ and BHT however, diffusion coefficients into HB307 are slightly higher than those into corn oil. In all the cases studied, the change in diffusion coefficient due to a change from corn oil to HB307 is less than a factor of 1.5.

The activation energies of the oligomers migrating from branched polyethylene into either corn oil or HB307 are in better agreement than the diffusion coefficients themselves as indicated in Table 3. The diffusion coefficients of $n-C_{32}H_{66}$ follow the same temperature variation regardless of whether corn oil or HB307 is used as the solvent. The extraction of $n-C_{32}H_{66}$ by trioctanoin also follows the same course. This is illustrated in Figure 3 for the extraction of branched polyethylene (sample 25) by the three different triglycerides. The activation energies for BHT and $n-C_{18}H_{38}$ extracted by corn oil and HB307 are also quite similar.

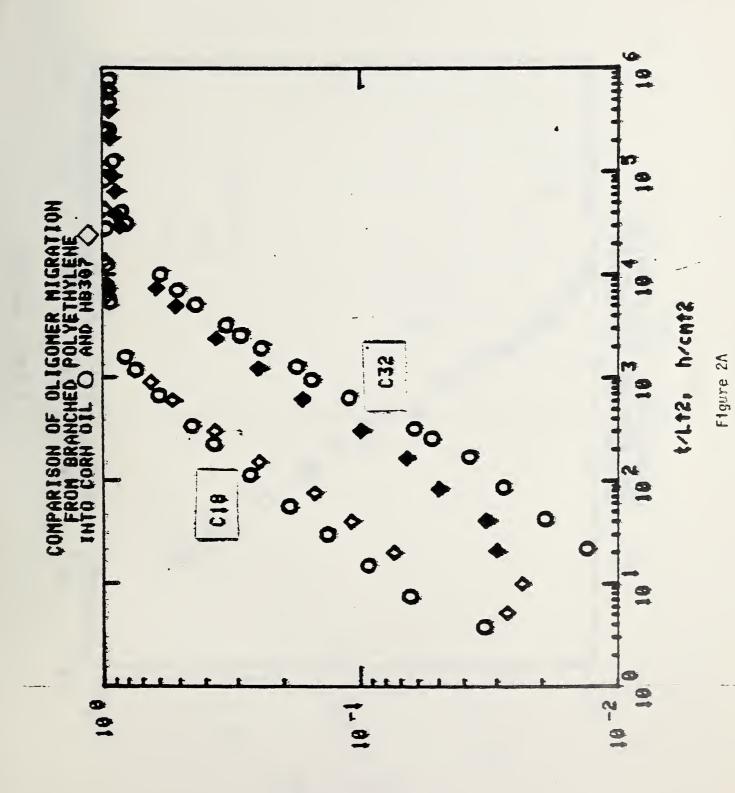
It may be concluded that the diffusion coefficients determined by extracting with all liquid triglycerides are identical well within a factor of 1.5 and the activation energies are probably within a factor of 1.3.

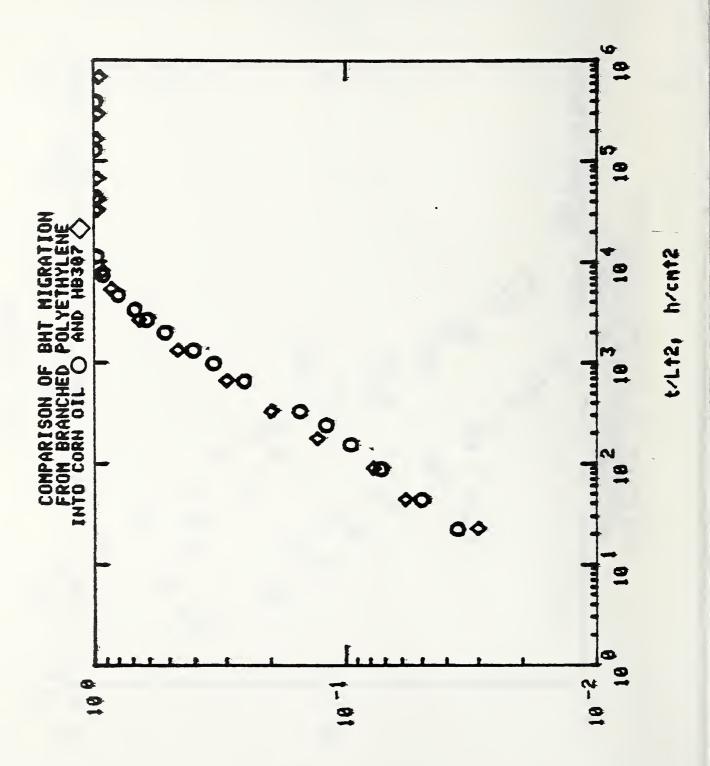
Table 2
Diffusion Coefficients of Migrants from Branched Polyethylene
0, µm²s⁻¹

	Polymer Code Migrant	9 1% n-C ₁₈ H ₃₈	16 1% n-C ₃₂ H ₆₆	25 45 ppm BHT
Solvent	T°C	, , ,		. F
Corn Oil	30 .	0.17	0.00034	0.029
HB307	40	0.18	0.0037	0.14
Corn Oil	60	3.4	0.20	0.83
HB307	60	2.5	0.29	0.96

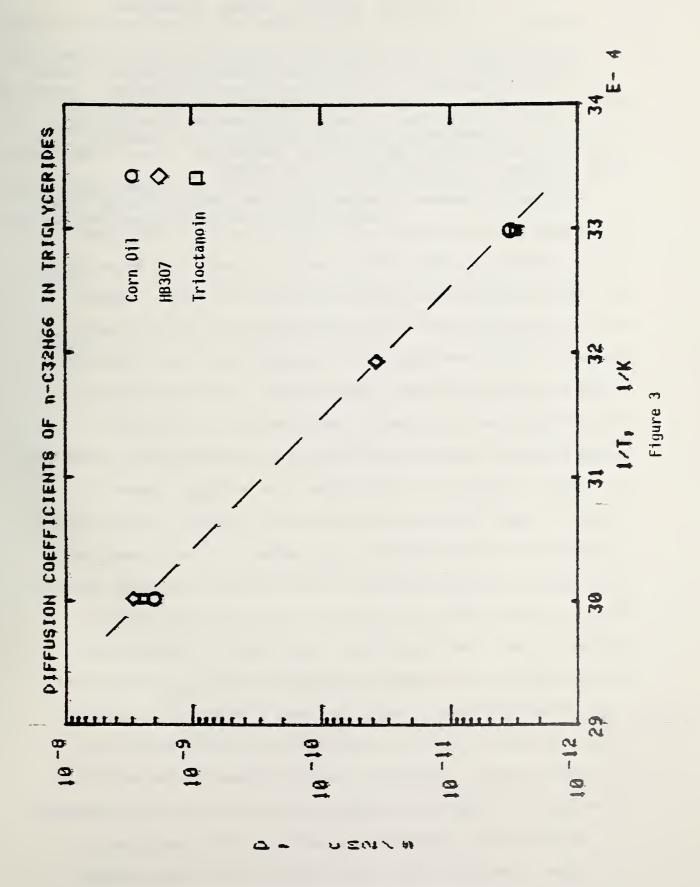
Table 3 Activation Energies for Migration from Branched Polyethylene $E_{\rm act}, k \, {\rm Jmol}^{-1}$

Polymer Code Migrant Solvent	9 1% n-C ₁₈ H ₃₈	16 1% n-C ₃₂ H ₆₆	25 45ppm 8
Corn Oil	84	180	94
нв307	114	190	84





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C. Effect of 5% Water on Ethanol Extractions

Anhydrous ethanol left exposed to the atmosphere will absorb moisture and eventually reach a constant boiling composition of 95% ethanol and 5% water. It is important to assess the influence of this amount of water in ethanol on additive migration into ethanol.

The results presented in Tables 4 and 5 and shown in Figures 4 and 5 indicate that the diffusion coefficients and partition coefficients for the migration of lower molecular weight oligomers, such as $n-C_{18}H_{38}$, are little affected by the presence of 5% water in ethanol. However, for $n-C_{32}H_{66}$ and BHT the diffusion coefficients are reduced by nearly one-half. This is consistent with the general conclusions in Section VI of the previous annual report, NBSIR 81-2264. Correlations developed there indicate that an increase in the extracting solvent polarity causes a greater reduction in the diffusivity of migrants with a relatively low diffusion coefficient in the polymer (i.e. n-C₃₂H₆₆) compared to those with higher diffusion coefficients (i.e. $n-C_{18}H_{38}$). The partition coefficients are also modified by the presence of 5% water, hence the total level of migrant extractable at equilibrium will be rather different. This is a direct result of the change in the solubilities of these migrants in the solvents considered. Such solubility changes can be rather substantial, for example the solubility of n-C₁₈H₃₈ in ethanolwater mixtures decreases by about six orders of magnitude as the water content increases from zero to 100% (Figure E5 of NBSIR 80-1999, the FY79 Annual Report). The effect of water in ethanol on the solubility of $n-C_{32}H_{66}$, an extremely hydrophobic migrant, may be even more pronounced.

We are currently studying the solubility of the three migrants in 95% ethanol. The exact value of the solubility of BHT may be somewhat difficult to assess since BHT degrades in solution over the extended

period of time required for the determination. Nevertheless, we hope to gain information about the relative solubility of BHT in anhydrous and 95% ethanol. The partition coefficients for BHT in 95% ethanol are simply an indication of the possible existence of partitioning rather than true values due to the combined effects of BHT decomposition during the extraction process, non-uniform distribution of BHT, and BHT grafted to the polymer sample during the molding process.

Table 4 Diffusion Coefficients of Migrants from Branched Polyethylene into Anhydrous Ethanol and 95% Ethanol D, $\mu m^2 s^{-1}$

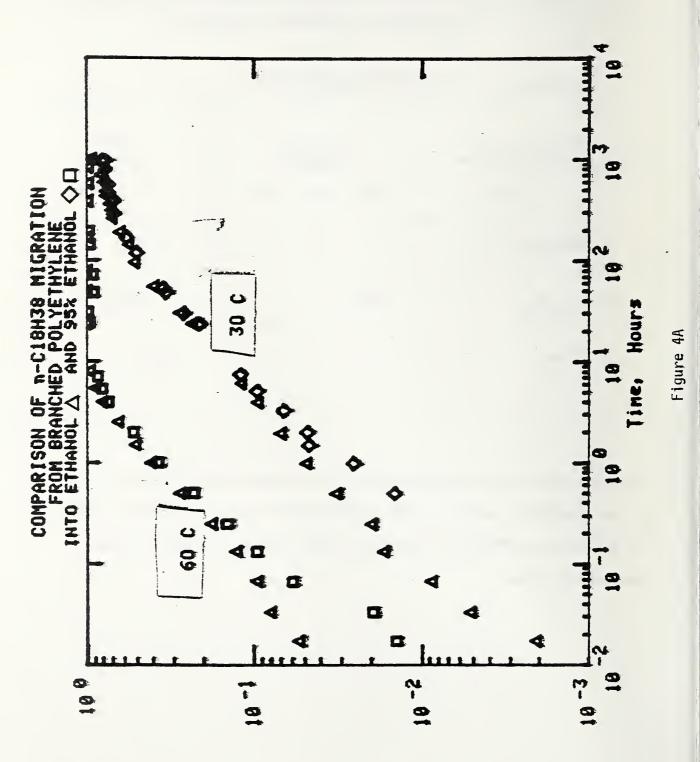
		Anhydrous		95% Eth	
Sample Code	Migrant	30°C	60°C	30°C	60°C
9	C18	0.061		0.050	
			3.0		2.5
16	C32	0.00015		0.000089	
			0.29		0.16
25	внт	0.038		0.020	
23	5111	0.000	7.0	0.020	0.67
			1.0		0.67

Table 5
Partition Coefficients of Migrants between Anhydrous Ethanol or 95% Ethanol and Branched Polyethylene

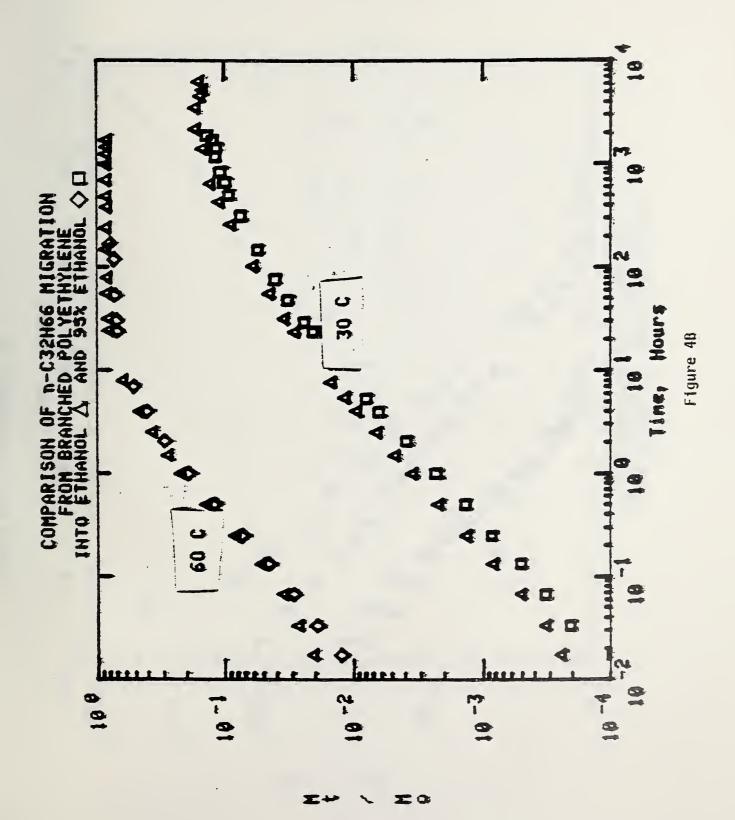
Partition Coefficient (k = Cs/Cp)

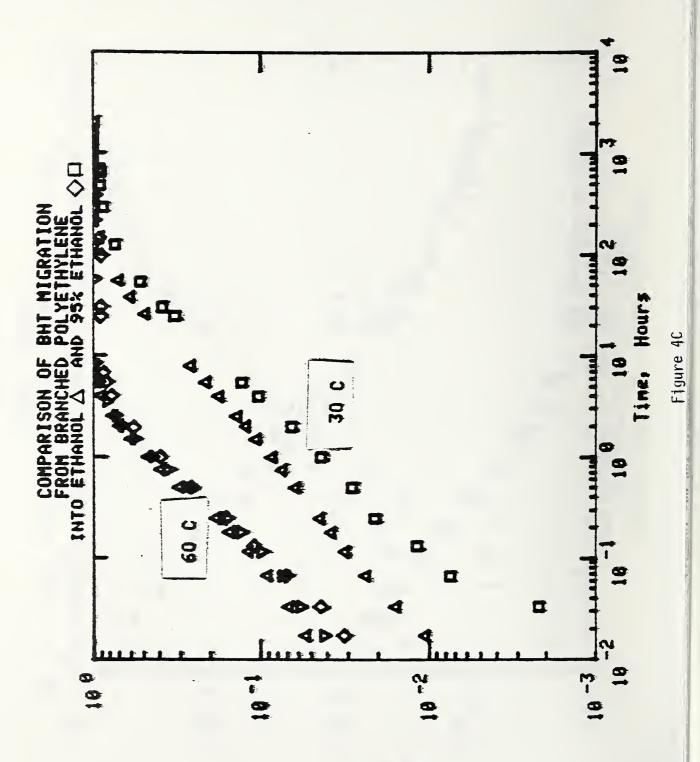
Migrant in Branched Polyethylene	Anhy <u>Eth</u>	95% Ethanol		
	30°C	60°C	30°C 60°C	
1% n-C ₁₈ H ₃₈	-	-	>0.045	
1% n-C ₃₂ H ₆₆	0.004	0.033	>0.00022	
0.01% BHT	-		~0.02	
		-	~0.02	

*For most extractions with 95% ethanol, the experiments have been carried to near completion but not quite to the point of equilibrium partitioning. The partition coefficient indicated in this column is therefore the lower-bound value.

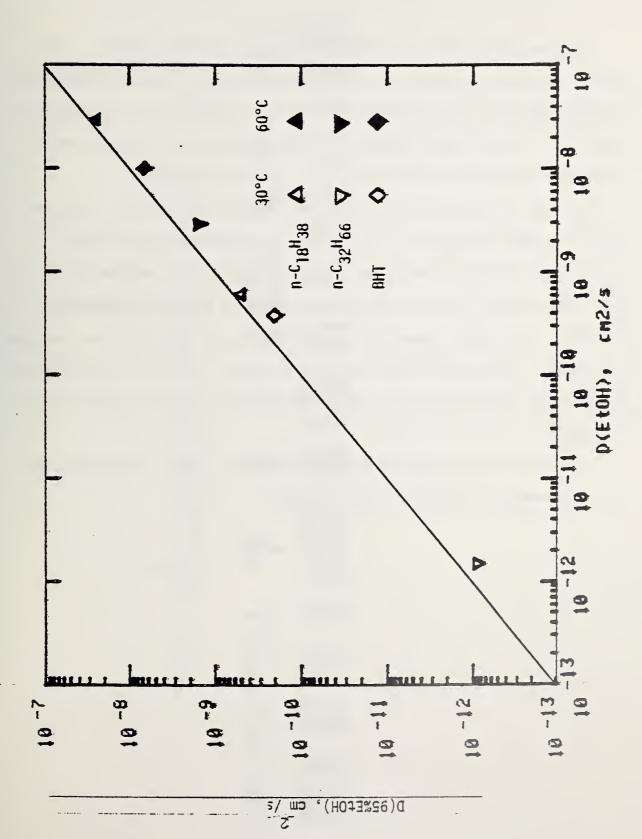


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II. Migration of BHT from Ethylene Vinyl Acetate Copolymers

Ethylene vinyl acetate copolymer of low vinyl acetate content is being used in increasing quantities as a food contact polymer. We are studying the migration behavior of the antioxidant BHT in this copolymer. The results are summarized in Table 6 and compared with the corresponding migration behavior in branched polyethylene in Figures 6A, B, and C and in Table 7.

BHT has a higher mobility in the (ethylene 5% vinyl acetate) copolymer than in branched polyethylene. The effect is more pronounced at lower values of the diffusion coefficient. For extractions by n-heptane there is only a 50% increase in the diffusion coefficient in E5%VA when compared to the BPE valve. Ethanol appears to be a stronger accelerating solvent compared to corn oil in the case of E5%VA (with an increase in the diffusion coefficient of about 50%) than for BPE (with an increase in the diffusion coefficient of about 20%).

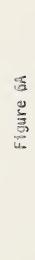
We are currently extending these experiments to lower temperatures and to a copolymer with 13% vinyl acetate content.

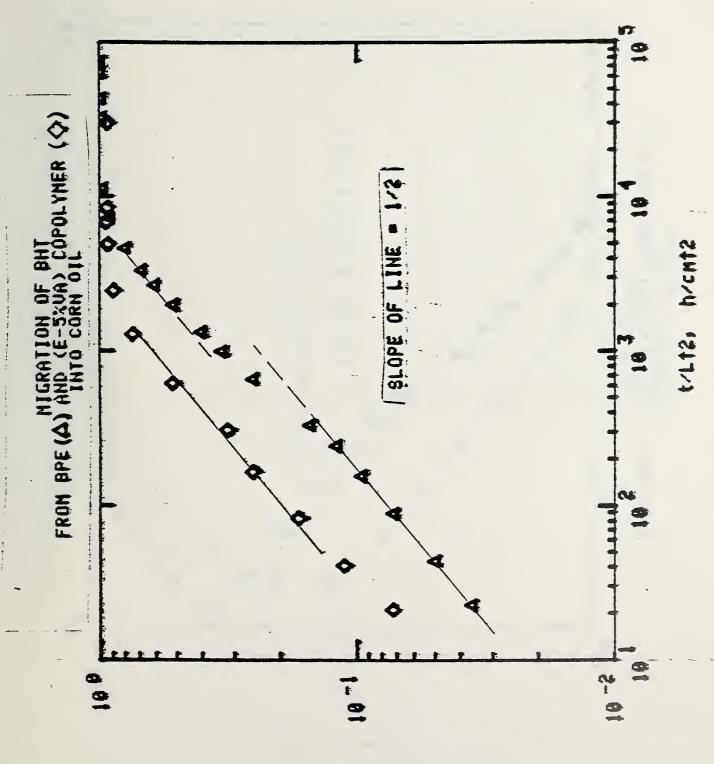
Table 6, Summary of Migration Data of BHT from Ethylene-5% Vinyl Acetate Copolymer

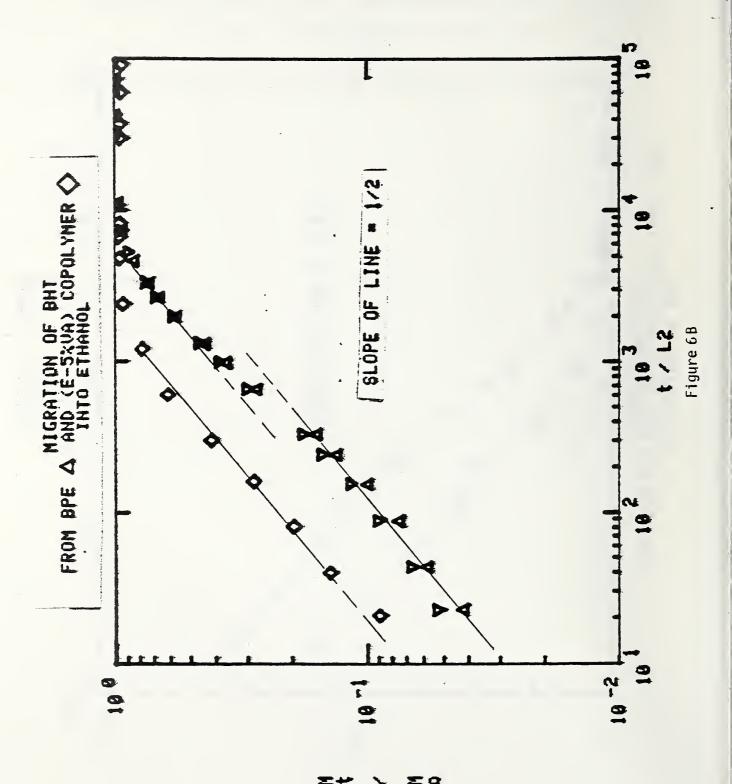
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Table 7 Comparison of Migration of BHT from Branched Polyethylene and from (Ethylene - 5% Vinyl Acetate) Copolymer at 60°C $_{D,\mu m}^{2} {}_{s}^{-1}$

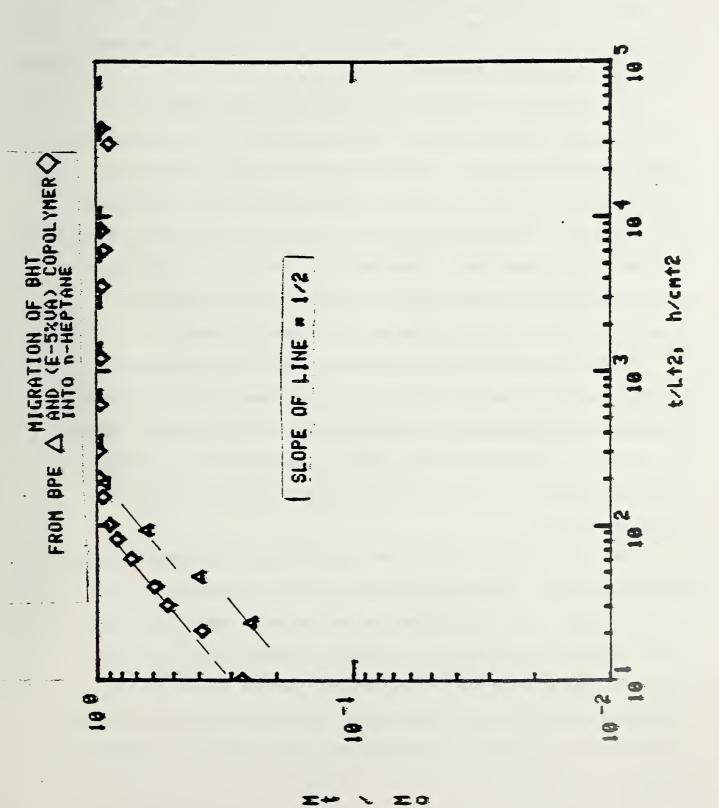
	BPE	E5%VA
Corn Oil	0.8	2.3
Ethanol	1.0	3.7
n-Heptane	37	57











III. Migration of VCM from PVC

The automatic head-space gas chromatograph, Perkin-Elmer Model F45, arrived in early April and is now awaiting set up and testing by the vendor before our acceptance is finalized.

Our first step will be to establish and calibrate the sensitivity of this instrument by using a series of standard solutions. The purported 2-3 order of magnitude increase in sensitivity of the electron capture detector (ECD) to VCM by the use of carrier gas containing up to 50ppm nitrous oxide will be studied. The ECD sensitivity as a function of nitrous oxide concentration is of major importance since a determination of the kinetics of VCM migration at very low levels is strongly tied to the stability of the sensitivity enhancement in the nitrous oxide concentration range of concern.

Extraction experiments may be performed on both powdered samples and molded plaques. The powdered sample may not yield good kinetic data because the sample is not monodisperse but has a particle size distribution. However, the submicron size particles may be useful for studying the equilibrium or partitioning properties that are very important in determining the final amount of migration.

Kinetic data on VCM migration from glassy PVC may be provided by studying the molded plaques. Relatively high temperatures are needed to mold rigid PVC and decomposition of the polymer can be detected at temperatures above 100°C. Although the main degradation product is hydrogen chloride, a trace amount of vinyl chloride may also be produced. We hope to overcome this problem by analyzing the total VCM content in the plaque before the extraction experiment and by studying or minimizing the effect of HCl on the determination of VCM.

We expect to have completed sufficient preliminary studies in this fiscal year to enable us to assess the reliability of the low level VCM migration studies.

IV. Review of the Migration of Organotin Compounds from PVC

Incorporation of a heat stabilizing material into poly(vinyl chloride) is necessary in order to limit thermal degradation and prevent discoloration at the high temperatures at which the PVC melt must be processed. These stabilizers are thought to react with HCl evolved from the polymer and prevent it from causing further degradation. The chemical reaction mechanisms underlying the stabilizing ability of the organotins in PVC have been reviewed in detail elsewhere (Ayrey and Poller, 1980). Another recent review on the performance of tin stabilizers under actual processing conditions is available (Cooray and Scott, 1980).

Two organotin stabilizers have been approved by FDA (Food and Food Products Regulations 121.2602, 1/20/68 and 178.2650, 1/16/75) for food packaging applications, di-n-octyltin-bis(2-ethylhexyl thioglycolate) (I), and di-noctyltin maleate polymer (II). These stabilizers, but primarily I, have been the subjects of numerous migration studies, with both foods and food simulants used as the extracting medium for stabilized, rigid PVC. Long term migration studies of I from PVC bottles into liquid foods have been conducted (Carr, 1969) and the concentration extracted after two months at 30°C given. The quantity of tin extracted from the bottles as organotin was often found to be comparable to the amount of "natural tin" or the background level of tin in the foods before extraction, indicating the importance of taking this factor into account to correct experimental total tin determinations. After six months, the amount of I migrating from PVC bottles into sunflower oil was found to be less than or equal to 2 ppm (Woggon et al., 1969). Beer stored in PVC bottles at 20°C for eight months contained 0.01 mg I/1 (Koch and Figge, 1971). Organotin stabilizers were readily extracted from PVC food containers by olive oil, orange or lemon soft drink concentrate, and aqueous ethanol (Brighton, 1968). Migration of I and II into a variety of liquid

foods has been studied at 57° C for eight weeks (Pinkasavage, 1970). Levels of organotins in these materials ranged from $0.01 - 0.24 \,\mu g$ organotin/g food. No additional changes in the level of migration were found when the study was extended at room temperature to a full year in duration.

Several studies have been conducted with food simulating solvents as the extracting medium. A brief review on the use of food simulating solvents in determining migration of dioctyltin stabilizers from PVC is available (Keller et al., 1975). Impact resistant PVC containing about 1% I was extracted with seven simulants (Dostal, 1980). The stabilizer was eluted over the range $0.004 - 0.045 \text{ mg/dm}^2$ polymer, the latter value for extraction with heptane. Studies of extraction kinetics comparing liquid foods to their corresponding simulating solvents have been reported (Keller et al., 1974). The total amount of stabilizer extracted varied from 0.06 µg I/g for type II foods to 0.44 μg I/g for type V foods. Stabilizer I could be readily detected in heptane and other organic solvents used to extract rigid PVC but could not be detected in coconut oil, triacetin, or triglyceride mixtures (Bergner and Berg, 1972). Radiolabeled stabilizers have also been studied. A determination of the total amount of all components migrating, so-called "global migration", has been made for rigid PVC stabilized with I (Figge, 1976). The fat simulant HB307-14C was used in these extractions. The total quantity of migrants extracted is equal to the weight lost by the polymer during storage in the simulant after correcting for the amount of simulant absorbed. The radiolabeled stabilizers methyltin-tri- (III), dimethyltin-di- (IV), and trimethyltinthioglycolic acid-2-ethylhexyl ester (V), 14C-labeled in both the methyltin or thioglycolate moiety, have been extracted with HB307 (Figge, 1975). The total amount of intact stabilizer migrating into fat-containing foods was found to be <1 ng III/g, 73 ng IV/g, and 39 ng V/g.

Typical food-grade PVC formulations include not only a stabilizer but also processing aids, lubricants, and colorants. Impact modifiers and plasticizers are often added. These materials can each be present at levels of 1-10% by

weight of the total molding compound. The influence of these additional components on stabilizer migration has been studied by several investigators. The chemical nature of organotin stabilizers was reported to have little effect on their migration from compounded PVC into water (Sheftel, 1978). The migration of dibutyltin-bis(isoctyl thioglycolate) (VI) stabilizer into water was found to increase as stabilizer and plasticizer (dioctyl phthalate) content increased. A study on the influence of epoxidized soybean oil, a lubricant, on the migration of dioctyltin and dibutyltin compounds into 50% ethanol, 5% acetic acid, and sunflower oil at 45°C for ten days concluded that lubricant concentrations below 3% have no effect on stabilizer migration (Woggon et al., 1967). Global migration from rigid PVC into water, 10% aqueous ethanol, and 3% acetic acid was found to be unaffected by the presence of up to 11% of this lubricant or by the addition of I (Thomas, 1980).

Detection of tin in foods is typically accomplished by oxidative destruction of the food with a nitric and sulfuric acid mixture followed by addition of HBr and HCl to form a tin halide which is then distilled and its concentration determined quantitatively by colorimetry. The distillation step can be omitted when food simulants are used. The method is sensitive to 5-7 µg tin, depending on whether dithiol or pyrocatechol violet is used as the reagent in colorimetric determinations. An anodic stripping method for determination of organotin compounds has also been described and an analytical scheme proposed for detection of these compounds and their decomposition products migrating from PVC (Woggon, et al. 1972). Radiolabeled stabilizer compounds have been used successfully as a sensitive method for quantifying migration, as discussed previously. Flameless atomic absorption spectroscopy (AA) has also been used in a recent study of organotins extracted from PVC by water and edible oils (Taleb et al., 1979). The water extracts were injected directly while the oil samples required dilution with cyclohexane. The limit of detection was reported to be 1.8 µg Sn/g in PVC and approximately 10 ng Sn/g in the simulants.

AA has been employed to detect octyltins extracted by water from PVC pipe (Dietz et al., 1979). Their quantitative analysis of tin was complicated by adsorption of extracted tin and precipitation of insoluble tin compounds. The procedure was modified by acidifying the solutions with concentrated HCl before AA analysis, which increased the tin level in solution by a factor of eight.

There is a lack of a consensus in the organotin migration literature on two issues: whether the whole stabilizers or their degradation products migrate into simulating solvents and the source of the stabilizer available for extraction. On the question of stabilizer degradation, the undegraded organotin compound was found to migrate from PVC into hexane and lipid based extractants while the major component migrating into aqueous extractants was an organotin oxide in the form of the cyclic trimer, $(R_2Sn0)_3$, with minor amounts of the undegraded compound and an alcohol form, $R_2Sn(OH)_2$, also thought to be present (Yen, 1970). Migration of di-n-octyltin esters and other organotins into aqueous and fatty foods has been studied (Woggon, et al. 1971). No chemical change in the octyltins was evident while for the other organotins studied mainly decomposition products were found to have been extracted. Experiments with I, radiolabeled at the ester or n-octyl group, migrating into edible oils showed that the stabilizer migrates either undegraded from rigid PVC or, following its hydrolysis, into two moieties which migrate at the same rate (Seidler et. al., 1971). The same material similarly radiolabeled has been the object of another study involving migration into edible oils and HB307 (Figge and Zeman, 1973). They concluded that the stabilizer and its decomposition products di-n-octyltin dichloride (VII) and oxide (VIII), 2-ethylhexyl thioglycolate (IX), and their decomposition products all migrate from PVC into fat-containing foods. In addition, free VIII was thought to form cyclic trimer and migrate into aqueous media. Further studies with this radiolabeled stabilizer (Figge et al., 1979) showed that it did not

migrate intact but VII and bis(2-ethylhexyloxycarbonylmethyl) disulfide (X), the dimer of IX, could be found in small amounts in aqueous or fat simulants. The quantities migrating (in $\mu g/dm^2$ polymer) for ten days at 40°C were given as follows: 0.04-0.17 VII and 0.14-0.47 X into water and 3% acetic acid, 0.50-2.12 VII and 0.51-0.99 X into HB307 and olive oil, and 0.88-5.48 VII and 3.63-3.81 X into heptane. If the stabilizers II, III, and IV discussed previously were transformed into the corresponding methyltin chlorides and thioglycolates before or during migration, their concentration in fatcontaining foods has been determined to be 0.5, 29, and 13 ng/g for the methyltin chlorides of II through IV, respectively (Figge, 1975). No definite conclusions can be drawn on the chemical nature of the migrating species of stabilizer I from the available literature.

Several studies present evidence that organotin stabilizers are extracted primarily from the surface of rigid PVC and not from bulk polymer as is normally assumed. A rapid initial migration into heptane and 95% ethanol followed by a period of further migration at a decreased rate was evident for samples containing 2% stabilizer (Downes and Gilbert, 1975, Downes, 1972). The initial amount of migration was greatly retarded when the PVC surface was cleaned with heptane prior to extraction. If the polymer was aged for two weeks following the surface cleaning, the retardation of migration was no longer observed upon extraction. A study on migration of decomposition products of I into edible oils and HB307 described previously concluded that since PVC cannot be swelled by these liquids, stabilizers within the polymer cannot migrate from the samples and only free surface stabilizer is available for extraction (Figge and Zeman, 1973). This point was amplified in a later study (Figge et al., 1979). Stabilizer I, some IX, and a trace of X could be found in the bulk polymer while only decomposition products VII and X could be identified on its surface. The latter two compounds were the species determined to have been extracted by the simulating solvents. These PVC

adsorption of extracted tin and precipitation of insoluble tin compounds. The procedure was modified by acidifying the solutions with concentrated HCI before AA analysis, which increased the tin level in solution by a factor of eight.

There is a lack of a consensus in the organotin migration literature on two issues: whether the whole stabilizers or their degradation products migrate into simulating solvents and the source of the stabilizer available for extraction. On the question of stabilizer degradation, the undegraded organotin compound was found to migrate from PVC into hexane and lipid based extractants while the major component migrating into aqueous extractants was an organotin oxide in the form of the cyclic trimer, (R2SnO)3, with minor amounts of the undegraded compound and an alcohol form, $R_2Sn(OH)_2$, also thought to be present (Yen, 1970). Migration of di-n-octyltin esters and other organotins into aqueous and fatty foods has been studied (Woggon, et al. 1971). No chemical change in the octyltins was evident while for the other organotins studied mainly decomposition products were found to have been extracted. Experiments with I, radiolabeled at the ester or n-octyl group, migrating into edible oils showed that the stabilizer migrates either undegraded from rigid PVC or, following its hydrolysis, into two moieties which migrate at the same rate (Seidler et. al., 1971). The same material similarly radiolabeled has been the object of another study involving migration into edible oils and HB307 (Figge and Zeman, 1973). They concluded that the stabilizer and its decomposition products di-n-octyltin dichloride (VII) and oxide (VIII), 2ethylhexyl thioglycolate (IX), and their decomposition products all migrate from PVC into fat-containing foods. In addition, free VIII was thought to form cyclic trimer and migrate into aqueous media. Further studies with this radiolabeled stabilizer (Figge et al., 1979) showed that it did not migrate intact but VII and bis(2-ethylhexyloxycarbonylmethyl)disulfide (X), the dimer

of IX, could be found in small amounts in aqueous or fat simulants. The quantities migrating (in $\mu g/dm^2$ polymer) for ten days at 40°C were given as follows: 0.04-0.17 VII and 0.14-0.47 X into water and 3% acetic acid, 0.50-2.12 VII and 0.51-0.99 X into HB307 and olive oil, and 0.88-5.48 VII and 3.63-3.81 X into heptane. If the stabilizers II, III, and IV discussed previously were transformed into the corresponding methyltin chlorides and thioglycolates before or during migration, their concentration in fatcontaining foods has been determined to be 0.5, 29, and 13 ng/g for the methyltin chlorides of II through IV, respectively (Figge, 1975). No definite conclusions can be drawn on the chemical nature of the migrating species of stabilizer I from the available literature.

Several studies present evidence that organotin stabilizers are extracted primarily from the surface of rigid PVC and not from bulk polymer as is normally assumed. A rapid initial migration into heptane and 95% ethanol followed by a period of further migration at a decreased rate was evident for samples containing 2% stabilizer (Downes and Gilbert, 1975, Downes, 1972). The initial amount of migration was greatly retarded when the PVC surface was cleaned with heptane prior to extraction. If the polymer was aged for two weeks following the surface cleaning, the retardation of migration was no longer observed upon extraction. A study on migration of decomposition products of I into edible oils and HB307 described previously concluded that since PVC cannot be swelled by these liquids, stabilizers within the polymer cannot migrate from the samples and only free surface stabilizer is available for extraction (Figge and Zeman, 1973). This point was amplified in a later study (Figge et al., 1979). Stabilizer I, some IX, and a trace of X could be found in the bulk polymer while only decomposition products VII and X could be identified on its surface. The latter two compounds were the species determined to have been extracted by the simulating solvents. These PVC

samples also showed a maximum amount of migration in the first aliquot of extracting solvent during long term extractions at 40°C with both HB307 and water. An additional complication could arise if some of the stabilizer, either on the surface or in the bulk material, is not free to migrate but has reacted with polymer molecules during melt processing of the material and become chemically bound to the PVC. This possibility has been mentioned previously but not thoroughly explored.

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Migration of three additives, n-octadecane, n-dotriacontane, and butylated hy-						
droxytoluene (BHT), from linear (LPE) and branched polyethylene (BPE) into several oil-						
simulating solvents is studied at 30 to 60°C. Diffusion coefficients of the three						
migrants from BPE into n-octanol are about the same as those into ethanol. The						
diffusivity of BHT	migrating from LPE i	nto n-octanol is two	to four times greater than			
that for ethanol extraction. Diffusion coefficients of the migrants from BPE into the						
			synthetic triglyceride HB307 are about equal to those in ethanol and other triglyceride			
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